Methods for Detecting and Measuring Ag Biotech Products



AEIC October 2004

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AEIC is pleased to provide the following slide presentation for use in educational or training applications associated with detection methods for biotech products. Due to the size of the file, this presentation is provided as a PDF, which does not allow for any changes in content. For a copy of the presentation on a CD please contact AEIC.

AEIC wishes to acknowledge the following companies and organizations for their contributions to the slides included in this presentation:

- EnviroLogix
- GeneScan
- Medallion Labs
- Monsanto
- SDI
- USFDA



Ag Biotech Crops

Transgenic plants have:

Novel trait (e.g., herbicide resistance) May express novel protein Novel DNA

• Novel DNA and protein may be found in:

Plant tissues Seed/Grain Food ingredients and food products



Biotech Crops 2004

• 2 major traits

Insect protection - *Bacillus thuringiensis* (Bt) Herbicide tolerance Roundup Ready (RUR) Liberty Link (LL) Bromoxynil tolerance (BXN)

• 4 major crops

Corn - Bt, RUR, LL Soy - RUR Canola - RUR, LL Cotton - Bt, RUR, BXN



Testing in Support of Labeling Biotech Foods

Consumer Choice

Approved Events – Quantitative and threshold testing

- European Food Labeling Law Labeling began April 10, 2000 and updated April 2004 Threshold adventitious < 0.9% - "genetically modified"
- Japanese Food Labeling Law Labeling began April, 2001 Threshold guidelines set at 5%

Unapproved Events - Detection

- Zero tolerance
 e.g. StarLink
- European Food Labeling Law Threshold adventitious < 0.5% - "genetically modified"



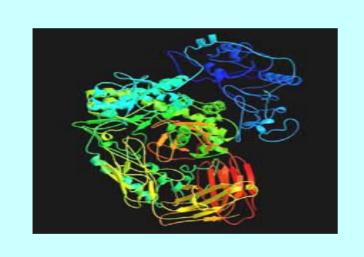
Determining Concentration of Biotech Ingredients in Foods

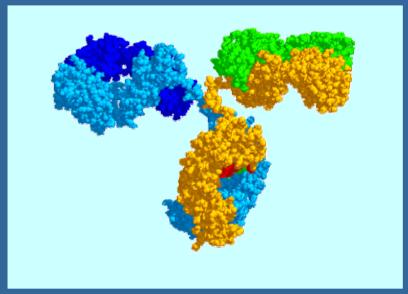
- Results are reported in terms of % Ag Biotech e.g. 1 Biotech corn kernel in 99 negative = 1%
- Decisions are based on regulated thresholds (given in weight %)
- Testing is based on detection/quantitation of novel DNA or protein
- Ag Biotech concentrations are estimated from protein concentration
- DNA can be measured in relative terms, i.e. % Roundup[®] Ready soybeans with respect to total soybean



Commonly Used Detection Methods

DNA-based methods PCR Protein-based methods Immunoassay (ELISA)







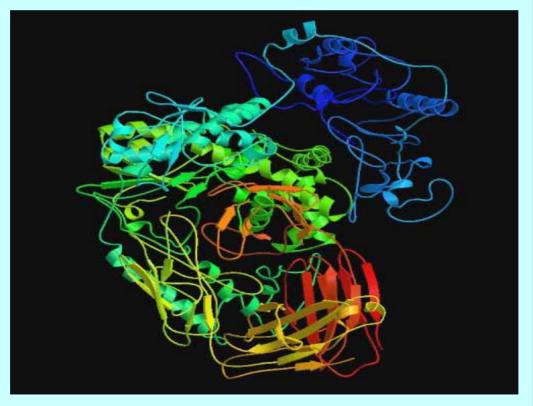
Biotech Protein - Expression and Detection

- Expression of new protein ranges from ng/g to μ g/g
- Cell contains many host proteins at very high concentrations
- Total protein methods not specific and not appropriate (e.g., Kjeldahl nitrogen, Near InfraRed)
- Method must be sensitive and specific



PCR - Polymerase Chain Reaction

Technique that allows amplification of specific DNA sequences millions of times in a few hours. DNA sequences unique to GM traits can be detected and measured.





PCR – Uses

- Genetic purity testing
- Disease Diagnosis
- Forensic Medicine
- Molecular Evolution
- Gene Cloning
- DNA sequencing

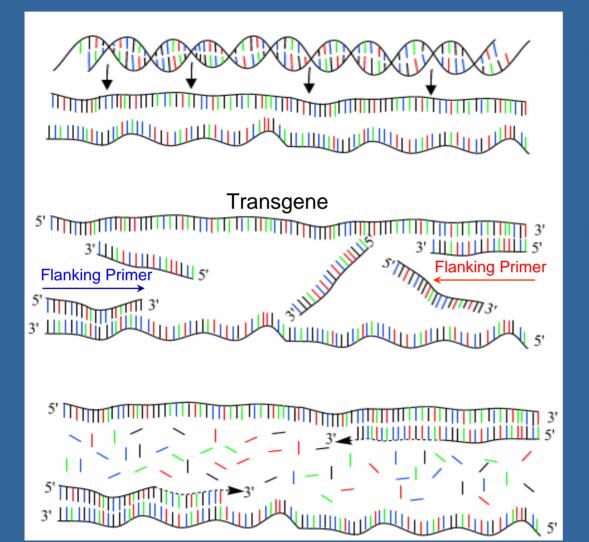




- High temperature is used to separate DNA Molecules into single strands
- Synthetic sequences of single-stranded DNA's (20-30 nucleotides) serve as primers that anneal to single-stranded target DNA molecules
- Two different primers are used to bracket a target region of DNA to be amplified: one strand is complementary to one DNA strand at one end of the target; the 2nd primer is complementary to the other end of the target region
- Primers are extended by thermal stable Taq polymerase
- Target DNA sequence is amplified
- Cycle is repeated 20-40 times



Polymerase Chain Reaction (PCR)



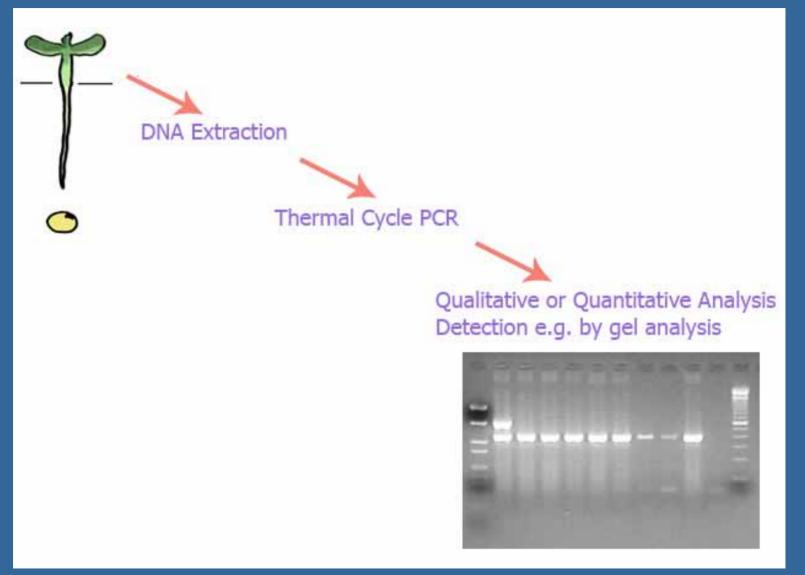
 Denaturation Step DNA denatures at high temperature (94C)

 Anneal Step Temperature lowered (50-65C) Primers anneal to target

Amplification Step
 Temperature raised (72C)
 Taq polymerase extends
 annealed primers
 Target DNA is amplified



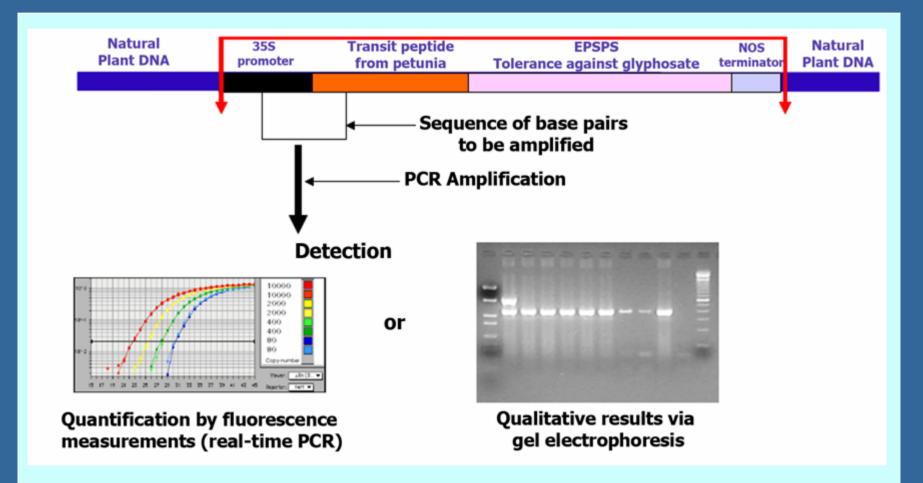
PCR Processing Steps





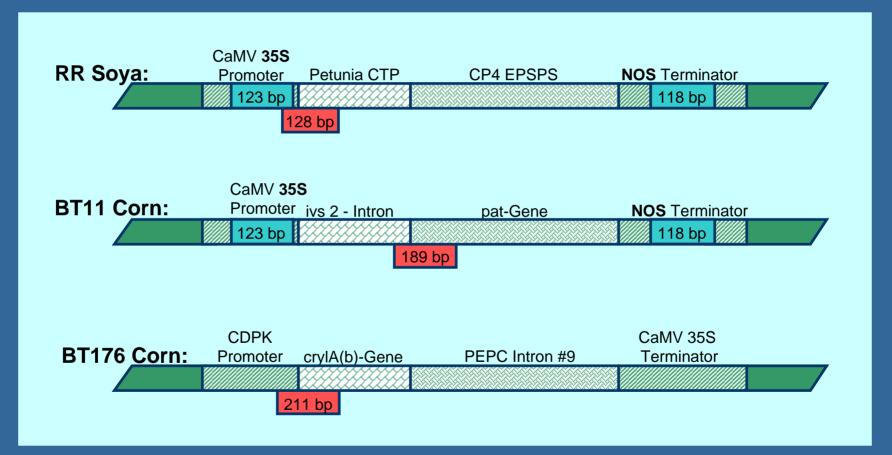
Has The DNA Been Modified?

Modified Genetic Sequence (RoundUp Ready[™]) Inserted Into Soy DNA





Some Synthetic Sequences





Applications of PCR

Qualitative

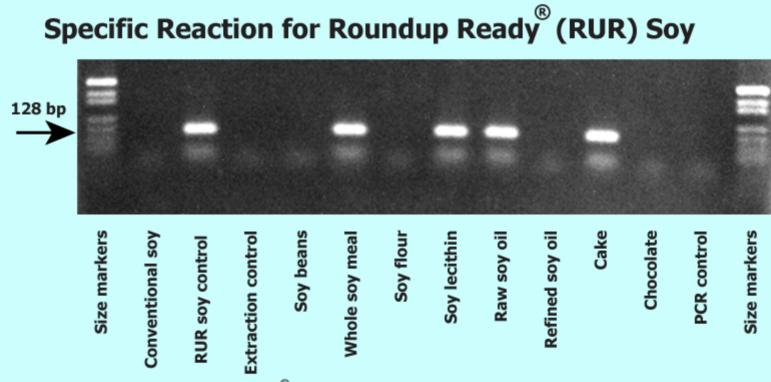
- A "YES" or "NO" answer
- Can look for specific DNA event such as RUR, MON810, etc. or
- Can look for generic elements such as NOS, 35S
- Applicable to a zero tolerance situation

Quantitative

- A determination of the percent of GM DNA present
- Relates amount of GM DNA to species DNA
- Real time quantitation by laser during PCR process
- Can be specific or a general screen



Qualitative Detection



Detection of Roundup Ready[®] soy-DNA (128 bp amplicon). The samples RUR soy. whole meal, lecithin, raw oil, and cakes contain genetically modified DNA



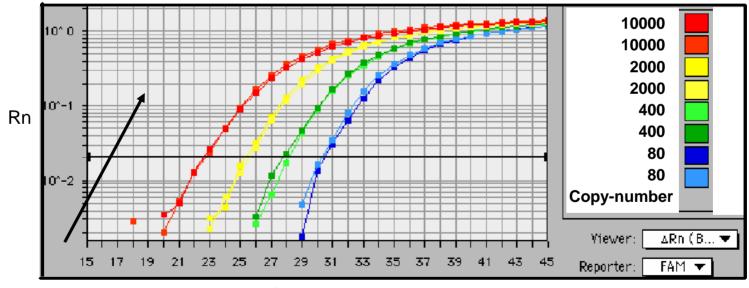
Quantitative Detection

- Real-time PCR measures the amount of PCR product at each and every amplification cycle
- Amplification plot is a curve that represents the accumulation of product over the duration of the entire PCR reaction
- A standard curve is generated that plots the cycle threshold values against starting amounts of DNA
- Measurement of DNA is done by fluorescing DNA molecules (TaqMan[™], Molecular Beacon, SYBR Green)



Quantification By Real-time PCR

Real-time PCR quantifies during the initial exponential phase. At higher number of cycles the fluorescence measurement reaches a plateau making a quantification impossible.



Cycle



PCR - Advantages

- High sensitivity
- Can detect and quantify specific traits
- Capable of detecting groups of traits through the use of common genetic elements (e.g. promotor or terminator)
- Higher stability of DNA (than of proteins) permits analysis of most mixed and processed foods



PCR - Disadvantages

- High cost per determination
- Requires sophisticated equipment and procedures
- Requires highly skilled and well trained personnel
- PCR reactions can be extremely sensitive to low levels of contaminating DNA template that will result in false positive reactions.
 - "Accidental PCR Template or amplicon carry-over"
 - (e.g. Reagents, pipettes, tips, fingers)
 - 10 pg of contaminating target DNA can produce 1 μg of product (25 cycles)
- Standardization across labs and protocols is still under discussion
- Labor intensive steps, needs time to complete (3 days or more)



False Positive Results

Problem	Control Measures
Artifact PCR products from templates other than the target sequence in question (worst scenario: artifacts of the same size as the expected PCR product)	 During validation: Search data base for potential unwanted (homologous) annealing sites Test new primer system on a wide range of DNA types that appear in food samples Verify the identity of the PCR product by Southern blot or restriction enzyme digestion
Corruption of sample, extracted sample DNA or PCR setup contains positive DNA	 During routine analysis: Follow strict rules how to work cleanly in a molecular biology lab Use one way sample flow lab design to prevent contamination with positive DNA from downstream steps (especially PCR products), strict separation of certain lab areas from each other Use duplicate analysis of each sample Use negative controls on homogenization, DNA extraction and PCR setup



False Negative Results

Problem	Control Measures
Insufficient sensitivity of primer system	During design and validation:
	Use computer aided primer design
	 Test the sensitivity of new primer systems for the intended concentration range
Inhibition of PCR reaction	During validation:
	 Test primer system on sample DNA solutions known to contain inhibitory compounds
	During routine analysis:
	 Test each PCR reaction for inhibition by an individual positive control (a spiked counterpart)
	Use duplicate analysis of each sample
DNA extraction failed	During design and validation:
	 Choose appropriate extraction protocol for sample type
	 Run controls to monitor efficiency of each extraction series



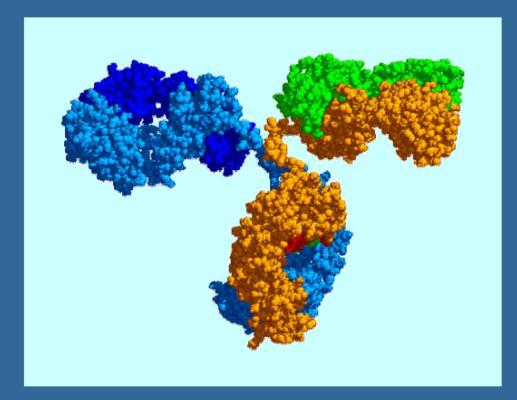
Protein Detection Methods

- Capillary Electrophoresis
- Chromatography
- Mass Spectrometry
- Immunoassay





An immunoassay is an analytical method which uses antibodies as reagents to quantitate specific analytes (proteins)



Tertiary Structure of an Antibody Molecule



Principles of Immunochemistry

- Antibodies physically bind target analytes
- Strength of binding determines sensitivity
- Specificity
 - Broad or specific (screening or quantitative)
 - Allows detection in complex matrix
 - Minimum sample preparation
 - Field-portable tests
- Sensitivity and specificity determined by antibody and assay conjugate pair

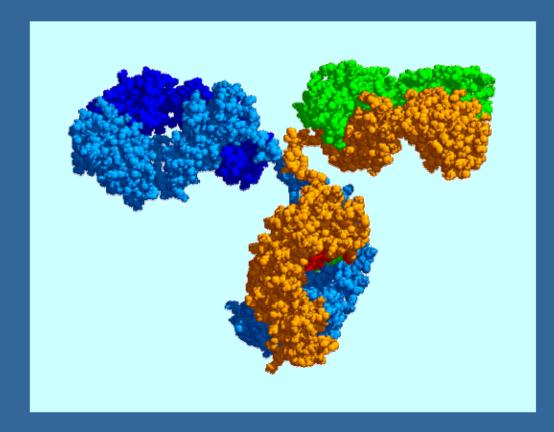


Immunoassays

- \$6 Billion Industry Worldwide
- 2.5 Billion Tests Sold Annually
- Highly Quantitative
- Recognized by Regulatory Agencies
- Flexible Test Formats
- Diverse Markets and Applications

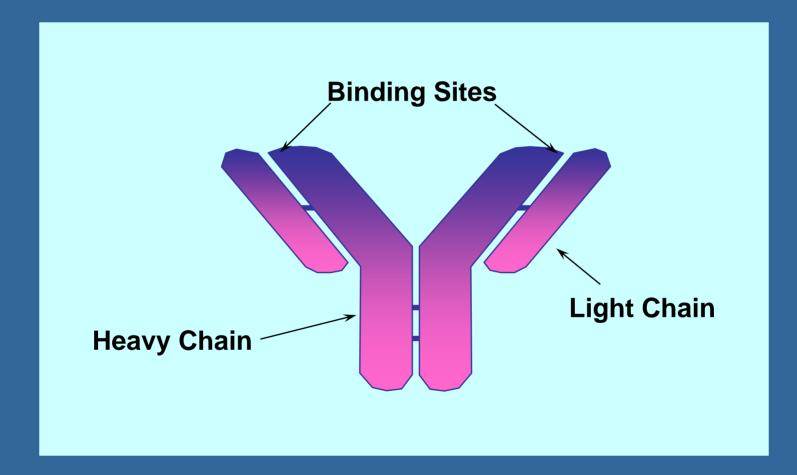


Antibody Structure





Antibody Structure





Immunoassay Formats

ELISA, Striptests, Western Blot

Each format has advantages and disadvantages

Fully automated – clinical analyzers Laboratory kits - ELISA Field tests – "Strip tests" 'Research' methods – Western blot

Choice of method is determined by specific application

Performance specifications
Ease-of-use (user training)
Testing location
Cost per test
Batch size, testing frequency
Turnaround time
Equipment costs



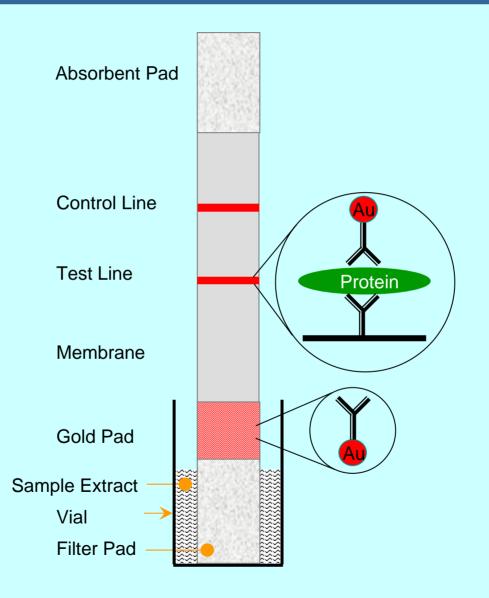
Lateral Flow Format



- 5 minute yes/no results
- Simple procedure for testing anywhere

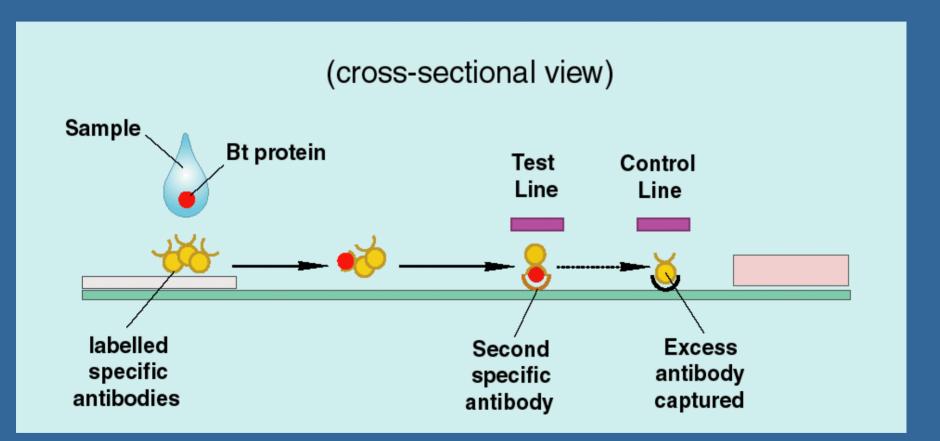


Strip Test



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Lateral Flow Format

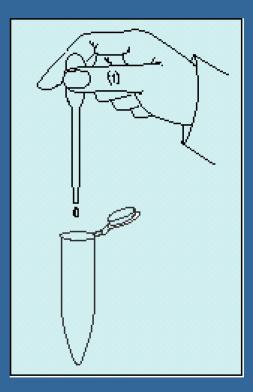




Lateral Flow Strips Testing Procedure

Extraction Step

- 1. Grind representative sample Add water (or buffer) & mix
- 2. Transfer extracted sample to vial





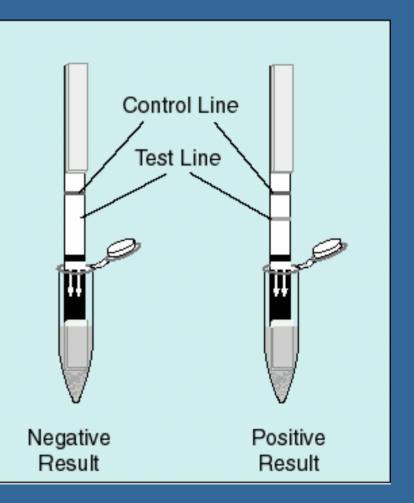
Lateral Flow Test Procedure

- 3. Insert strip into vial
- 4. Read Results

Negative = No test line, but control line visible

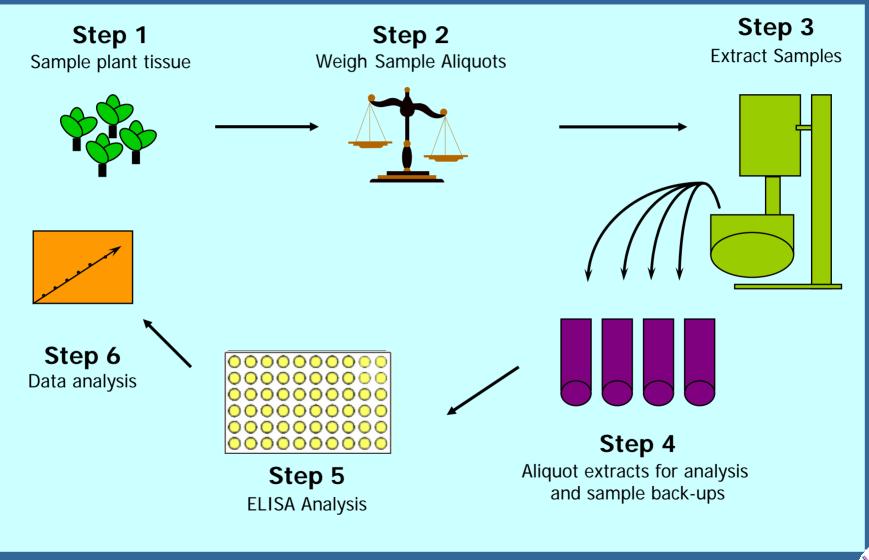
Positive = Visible test line and visible control line

Invalid test = No test line No control line





Steps in Determining Protein Expression by ELISA



Summary of Protein Testing by Immunoassay

- Immunoassays are quantitative analytical methods
- Flexible format provides for diverse applications
- Commercial methods are available for detection of biotech products

Raw agricultural commodities Processed food fractions

- Methods have been validated internationally
- Accurate, reliable and timely analysis



Immunoassay Development Process

• Define Performance Characteristics

- Sensitivity and specificity are determined by the nature and amount of antibody and the assay conjugate
- Format determined by application

• Development Process

- Antibody and assay conjugate design and development
- Test format and optimization
- Validation
- Controlled use of assay, QA/QC
- 1 to 2 Years



Immunoassay Performance Characteristics

- Sensitivity (LOD, LOQ) ppb to ppt (10⁻¹²M)
 LOD level of detection
 LOQ level of quantification
- Specificity
 - Families of chemicals vs. single compounds
 - Commercial products
 - Metabolites, degradation products
 - Process by-products, intermediates
- Precision repeatability, reproducibility
- Accuracy recovery and false negative/positive rates
- Matrix effects/interfering substances
- Quantitative range
- Stability, Reliability, Robustness
- Fitness for purpose



Assay Validation/Bridging for Key Tissues

- Sensitivity

 limit of detection, limit of quantitation

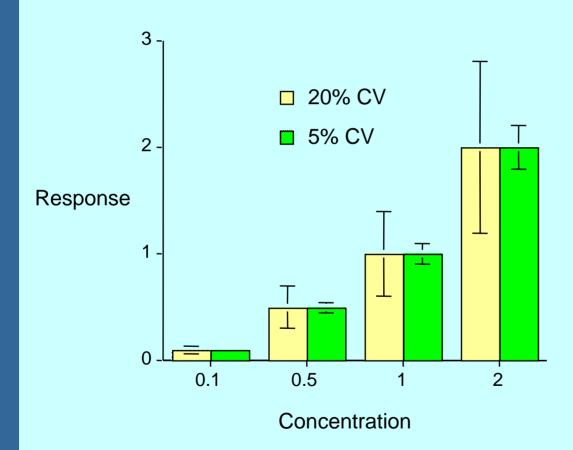


Precision

- The closeness of agreement between independent test results obtained under stipulated conditions
- The amount of variability due to random error
- Expressed as the standard deviation (sd) or coefficient of variation (cv)
- Determined by testing replicates of samples
- The precision of the final concentration must include variability of sample preparation
- Good precision provides ability to distinguish closely related concentrations
- Repeatability (reproducibility) is precision under repeatability (reproducibility) conditions
 - Repeatability conditions, test results obtained in the same lab, same method, same conditions, same operator, within short period of time
 - Reproducibility conditions, test results with the same method in different labs, different operators and different equipment



Effect of Precision on Ability to Discriminate Between Concentrations



Error bars for concentrations determined by the method having a 20% CV overlap. Both methods give the same concentration but it is not possible to say that adjacent concentrations differ from each other in the assay with the 20% CV within the given confidence interval.





- The closeness of agreement between the (or a) reported result and the accepted reference value
- Quantitative methods
 - Expressed as '% Recovery' of 'true' value
- Qualitative methods
 - Expressed as rate of false positive and negative results
- Influenced by
 - Precision Bias Matrix effects



Sample Extraction

• Matrix effects

- Sample matrix Extraction of the protein also extracts other substances from the sample
- Interfering substances Substances affecting assay performance

• Extraction efficiency

- The % of target protein extracted by the method protocol
- Does not need to be 100% must be consistent
- Balance
 - Sensitivity
 - Cost
 - Time
 - Ease-of-use



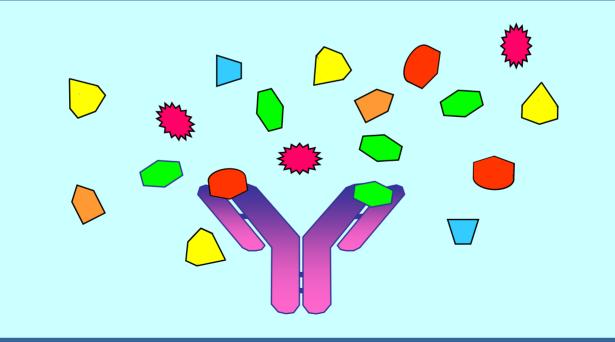
Sensitivity

- Change in the response divided by the corresponding change in the concentration of a standard (calibration) curve; i.e. the slope of the analytical calibration curve
- The minimum concentration of the target analyte (e.g. protein) that can be detected or quantified
 - Limit of detection (LOD) The concentration of protein which can just be detected by the assay (e.g. 50% of the time)
 - Limit of quantification (LOQ) The concentration of protein that can be quantified with stated precision (e.g. ±10%)
- Total method sensitivity must account for extraction efficiency



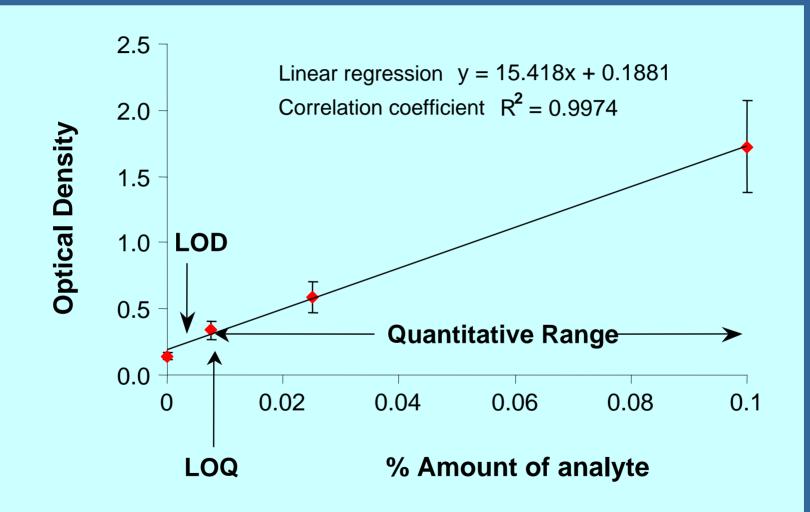
Specificity/Crossreactivity

- **Specificity:** the property of a method to respond exclusively to the characteristic or analyte (e.g. protein) defined in the method
- **Crossreactivity:** antibody binding at the binding site with substances other than the analytes of interest





Standard Curve





Standards and Controls

• Quantitative Methods

- A series of standards (calibrators) are used to construct a standard curve
 - Matrix-matched standards (e.g. corn flour)
 - Samples and standards are normalized for matrix effects and extraction efficiency
 - Must create standards for every matrix
 - Purified protein standards
 - Must establish that they give same response as protein in matrix

Qualitative methods

- Negative control
- Positive control e.g. threshold concentration, need to determine LOD
- May have multiple levels to group results in limited ranges (buckets)



Biotech Immunoassay Method Validations

• Collaborative studies

AACC MON810 Cry1Ab ELISA – ground corn StarLink Cry9c ELISA – corn flour and meal Joint Research Centre, European Union Roundup Ready[®] ELISA IRMM ground soybean certified reference materials Soy toasted meal, protein isolate, defatted flakes FDA

StarLink Cry9c ELISA – processed food fractions

USDA Certification

Cry9c strip tests – corn kernels Cry9c ELISA - ground corn, meal, flour CP4EPSPS Strip tests - ground corn, soybeans



Factors Effecting Use of Protein Immunoassays

- No protein no Immunoassay
- Very low level expression (e.g. Bt 176)
- Crossreactivity (e.g. GA21 Roundup[®] Ready corn)
 - Modified corn EPSPS 2 amino acids of 445 different from native corn EPSPS



Factors Effecting Quantitation Using Immunoassays

- Method performance characteristics (e.g. precision, accuracy)
- Variability of protein expression levels
 - Within an event
 - Between events expressing same protein (e.g. Cry1Ab)
- Varied effects of sample processing on protein conformation and antibody binding



Commercially Available Protein-based Tests For Commercial Ag Biotech Traits

Trait	Agdia	EnviroLogix	Neogen	SDI
CP4 EPSPS		E,S	S	E¹,S
Cry1Ab	E,S	E,S	E,S	E²,S
Cry9C		E ³ ,S	E,S	E ³ ,S
Cry2A		E,S		
Cry3A	E			
NK603		S	S	S
		1		

E ELISA

S strip

¹validated by JRC ring test for Roundup Ready Soy bean ²validated by AACC ring test for YieldGard[®](Mon810) Corn ³official method AACC international collaborative trial



Advantages of Immunoassay Methods

- Directly measure biologically active protein of interest
- Reliable
- Quantitative analysis
- Qualitative analysis
- High sample throughput
- Easy to perform and transfer to other laboratories
- Widely accepted method by regulatory agencies
- Established use in food industry
- Cost effective
- Timely Analysis



Disadvantages of Immunoassay Methods

- No single method will detect all biotechnology-derived products (DNA or protein-based)
- Methods are trait specific and must be validated for each matrix (DNA and Protein-based)
- Limited to use of protein containing processed ingredients and final food products
- Some products may not express a detectable protein in grain
- Antibodies may cross react



Monoclonal vs. Polyclonal Antibodies

Monoclonal

Polyclonal

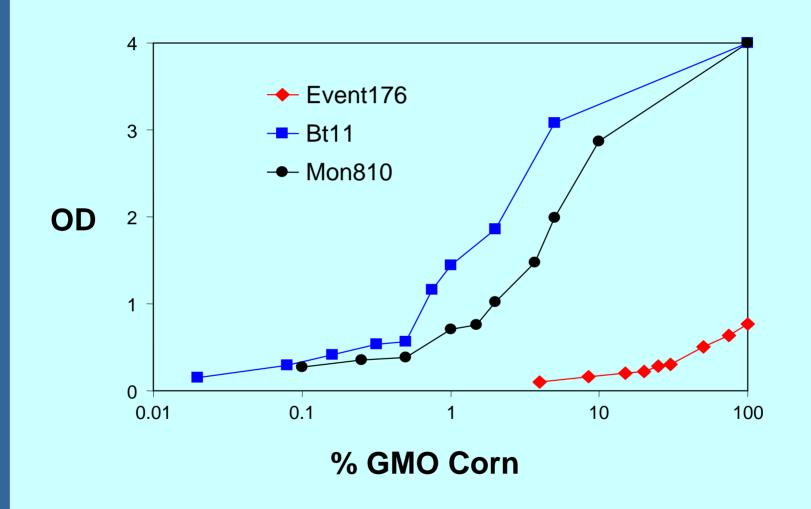
- Lot-to-lot consistency
- Indefinite supply
- Highly specific
- Longer lead time
- Higher initial costs

- Lot-to-lot variability
- More broadly reactive
- Often more sensitive
- Shorter lead times
- Lower initial costs

Selection is based on application, time and money

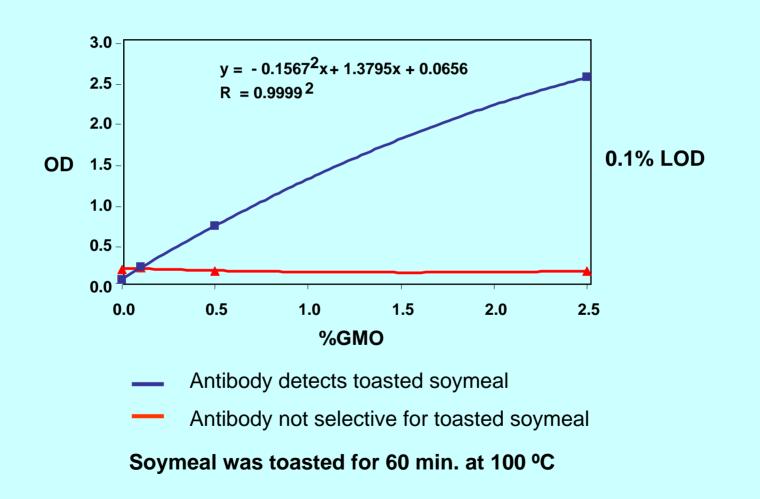


Reactivities of Different Varieties of Bt Corn in Cry1Ab ELISA





Reactivity of 2 Different ELISA to RUR Toasted Soy Meal





Clinical Diagnostic Immunoassays

- In Use >30 Years
- Basis for Critical Human Health Decisions
 - Disease diagnosis (AIDS, Hepatitis, PSA) Therapeutic drug monitoring Drug abuse screening Over 70 clinical analytes tested by immunoassay Home pregnancy tests
- Highly Reliable





Antibodies

- Key reagents in all immunoassays
- Proteins produced by immune system of higher animals

Produced by specific white blood cells In response to recognition of "foreign" substances Examples:

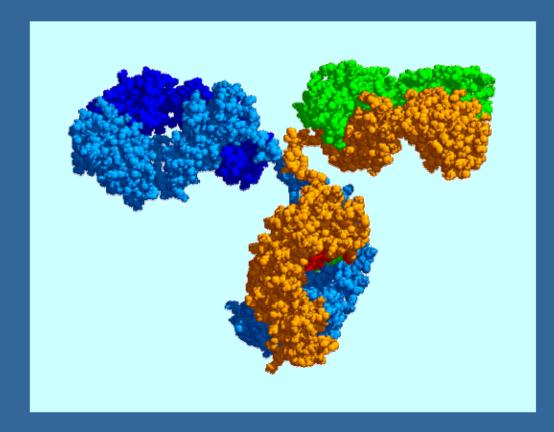
Vaccinations

Response to natural infections (mumps, chicken pox)

- Chemically bind to "Antigens"
- Tightly bind only to specific structure in substance which elicited production (Specificity)
- Strength of binding (Affinity) determines sensitivity of method
- Specificity allows detection in complex matrix minimum sample preparation

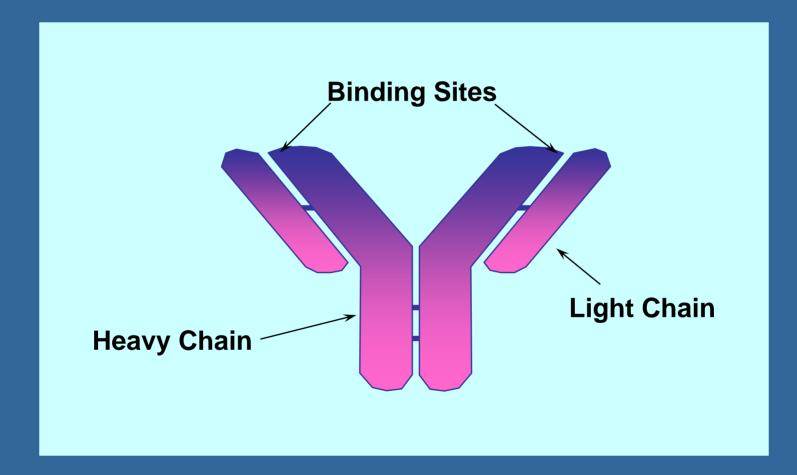


Antibody Structure



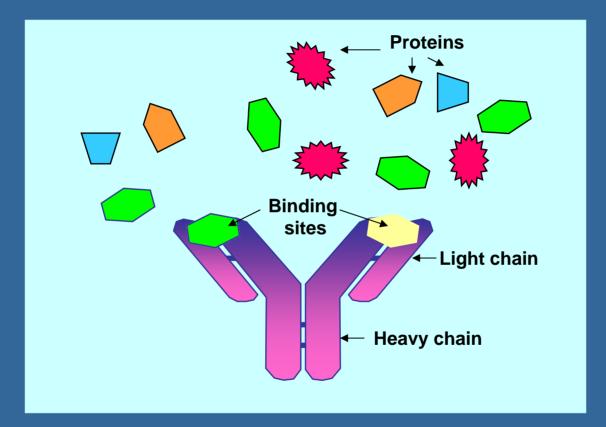


Antibody Structure





Antibody-Antigen Specificity



Antibodies only react with the epitope that elicited their production. Antigens must "fit" into the binding site.



Polyclonal Antibodies

- Polyclonal = "many cells"
- Animals (e.g. rabbits) are injected with target protein
- Many different antibody-producing cells make many different antibodies with many specificities
- Each antibody producing cell makes only one antibody with a single specificity
- Polyclonal antibodies purified directly from blood



Monoclonal Antibodies

- Monoclonal = "one cell"
- Mice are injected with the analytical target
- Antibody producing cells are taken from the animals
- Antibody-producing cells are fused with cells that grow continuously in culture to form "Hybridomas"
- A single hybridoma produces only one antibody
- A single hybridoma divides to produce a large population of 'clones' all making the same "Monoclonal" antibody
- Living hybridomas are frozen indefinitely in liquid nitrogen
- Indefinite supply of uniform consistency reagent



Immunoassay Reagents and Test Components

- Antibodies are purified and attached to a 'solid phase' to provide a means for separating the target protein from the sample
 - Plastic wells, tubes, capillaries
 - Membranes
 - Magnetic particles
- Antibodies are 'labeled' with detectable substances to provide a means for detecting and quantifying the presence of the target protein e.g.
 - Colored particles (e.g., colloidal gold, latex)
 - Enzymes
 - Fluorescent molecules
 - Chemiluminescent molecules
- Choice of format determined by application



Types of Immunoassays

- Competitive
- Double Antibody Sandwich



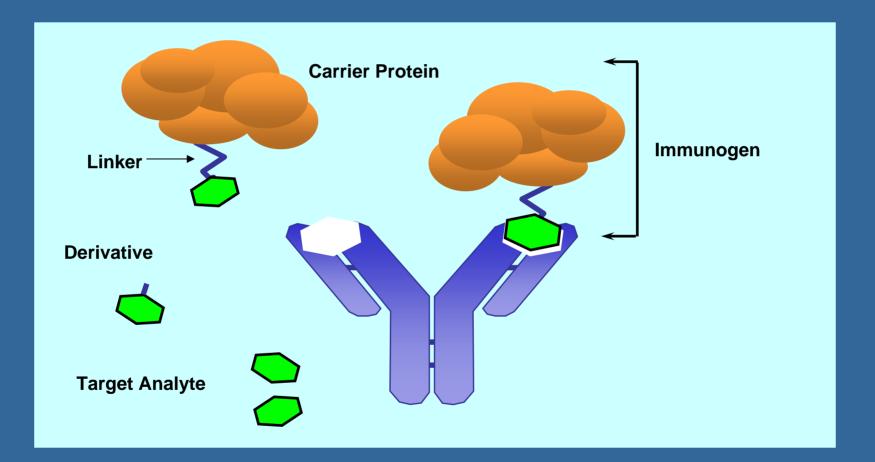
Antibody Development

- Immune system responds best to high molecular weight "Immunogens" M.W. Typically > 10,000
- Agrochemicals and environmental pollutants mostly small molecules M.W. Typically < 1,000
- Agrochemicals require preparation of suitable immunogen

Couple chemical to carrier protein

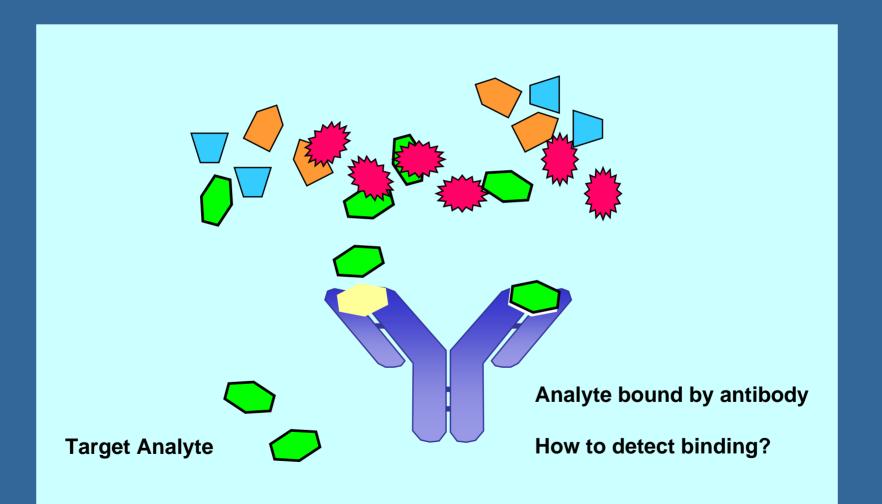


Immunogens



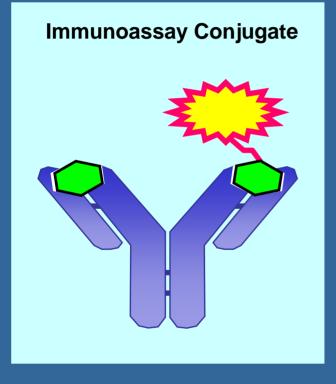


Immunoassay Visualization





Immunoassay Conjugates Detecting Binding

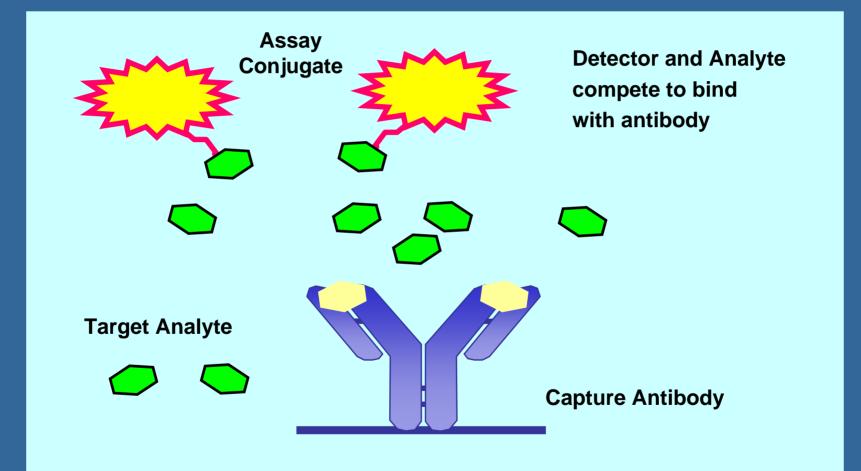


Detectable Label

Radiolabel (RIA) Enzyme (EIA) Fluorescence (FIA) Luminescence Electrochemical Visual Colloidal gold Colored latex



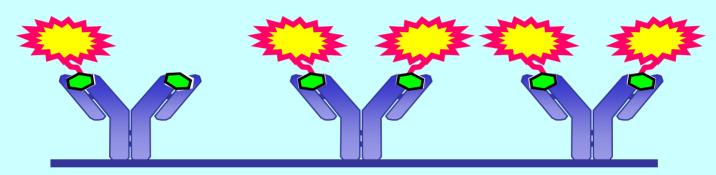
Competitive Immunoassay



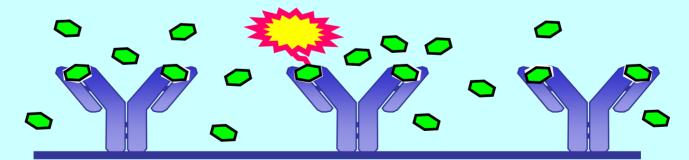


Competitive Immunoassay

I. No Analyte - high detection signal



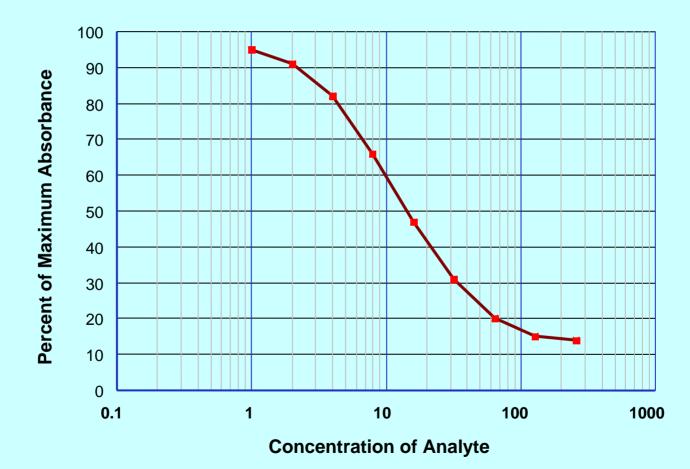
II. Analyte present - detection signal reduced





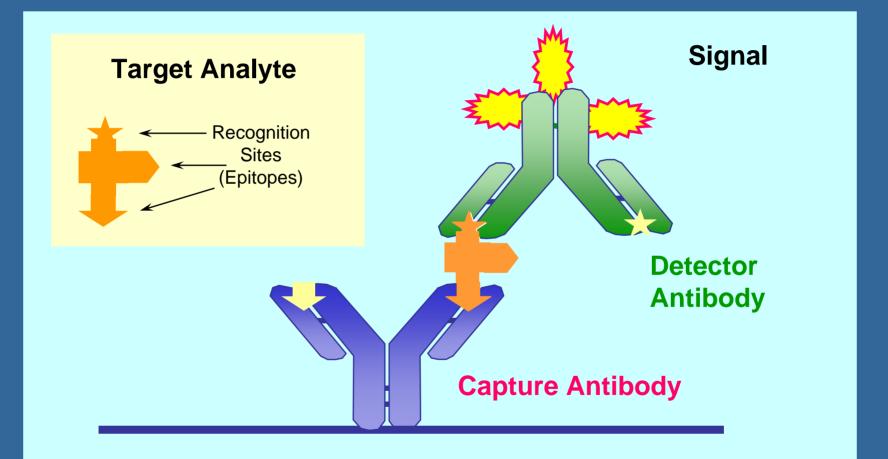
Competitive Immunoassay Data Format

Competitive Immunoassay Data



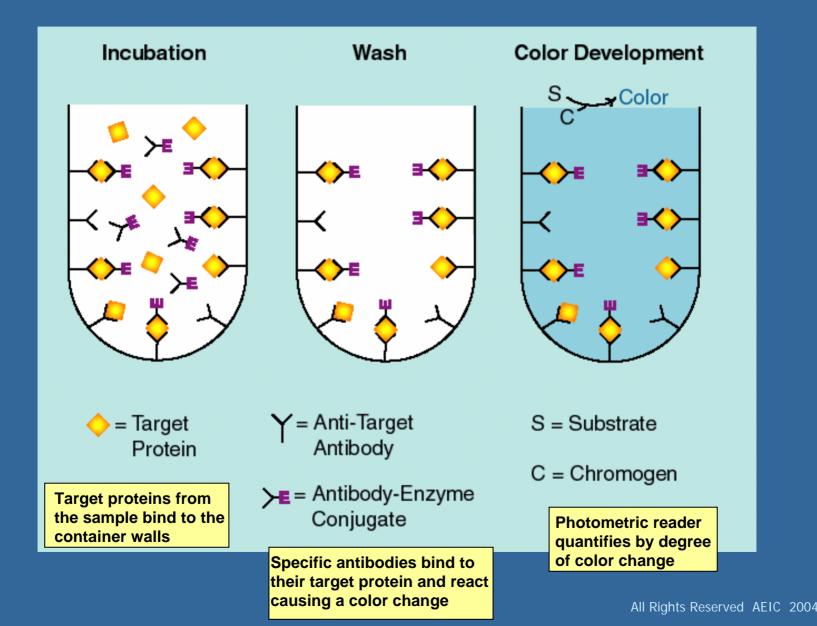
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Double Antibody Sandwich Immunoassay



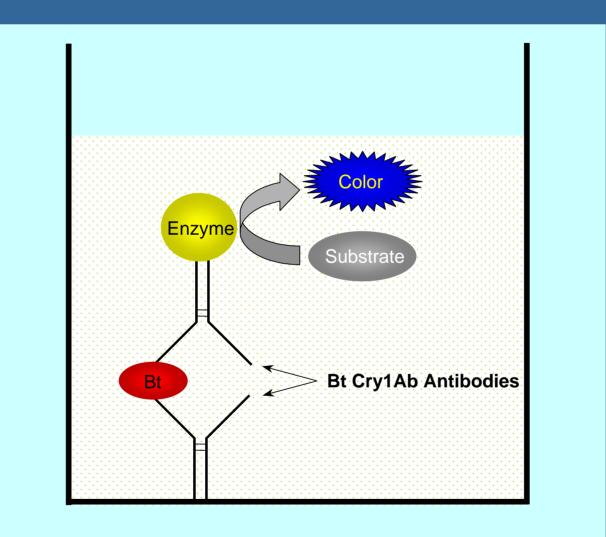


Sandwich Immunoassay Principle



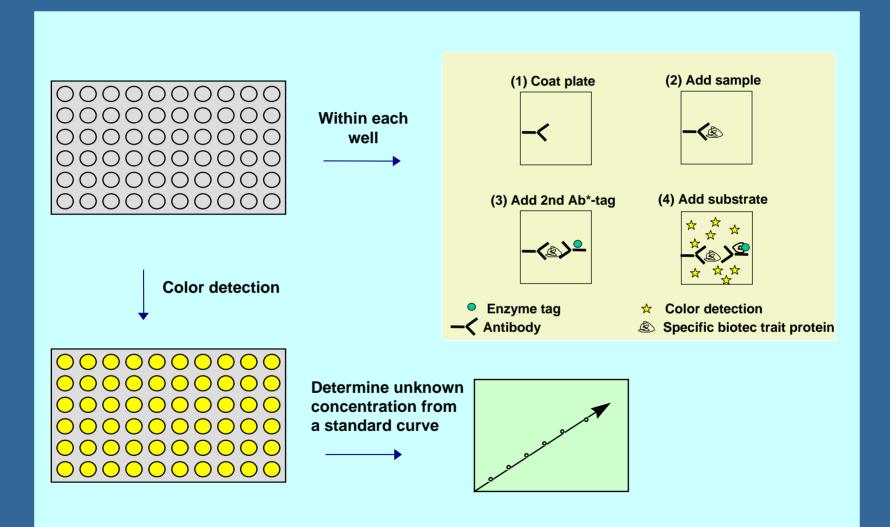
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Bt Cry1Ab Sandwich ELISA



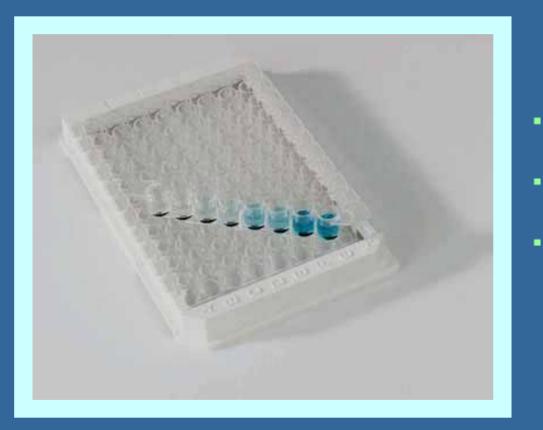


Protein Based Detection by ELISA





Microwell Plate Format



- Quantitative
- 1-2 hours
- Plate reader required



Coated Tube Format



- Qualititative or semi-quantitative
- Visually read color reactions (no equipment required)



Comparison of PCR, ELISA and Strip Methods

	PCR	ELISA	Strips
Design	DNA/Line specific	Protein specific	Protein specific
Requirements	Sophisticated	Moderate	Easy
Assay time	3-14 days	2-7 days	5-10 min
Sample cost	\$400-600	\$100+	\$7-10
Availability	Published/Testing Co	Diagnostic Co.	Diagnostic Co.
Validation	JRC (qualitative)	JRC/USDA	USDA
Application	Qualitative/semi-quant	Qual/semi/quant	Qual/ test/ compliance with threshold
Limitations	Basic pH, heat, false pos, false neg dynamic sys	heat, extraction, reference materials Protein dependent	Same as ELISA pro-zone, 1 sample/test
Sensitivity	│ ~>0.1% GMO	/~>0.3% GMO	~1% GMO



Other Immunoassay Markets

- Agricultural
- Environmental
- Veterinary
- Food
- Industrial
- Pharmaceutical
- Water Quality

- Microorganisms
- Allergens
- Hormones
- Toxins
- Pesticides
- Antibiotics
- Species identification
- Food adulteration
- Quality



Clinical vs. Environmental Immunoassay

The Sample

Clinical

Urine, blood, saliva



- Environmental, Agriculture (more diverse matrices)
 - Water
 - Soil extracts
 - Plant extracts
 - Animal products/tissues blood, urine, milk, meat
 - Food
 - Industrial processes and effluents







